

Isolation of a High Affinity Cannabinoid for Human CB1 Receptor from a Medicinal Cannabis Variety: Δ^9 -Tetrahydrocannabitol, the Butyl Homologue of Δ^9 -Tetrahydrocannabinol

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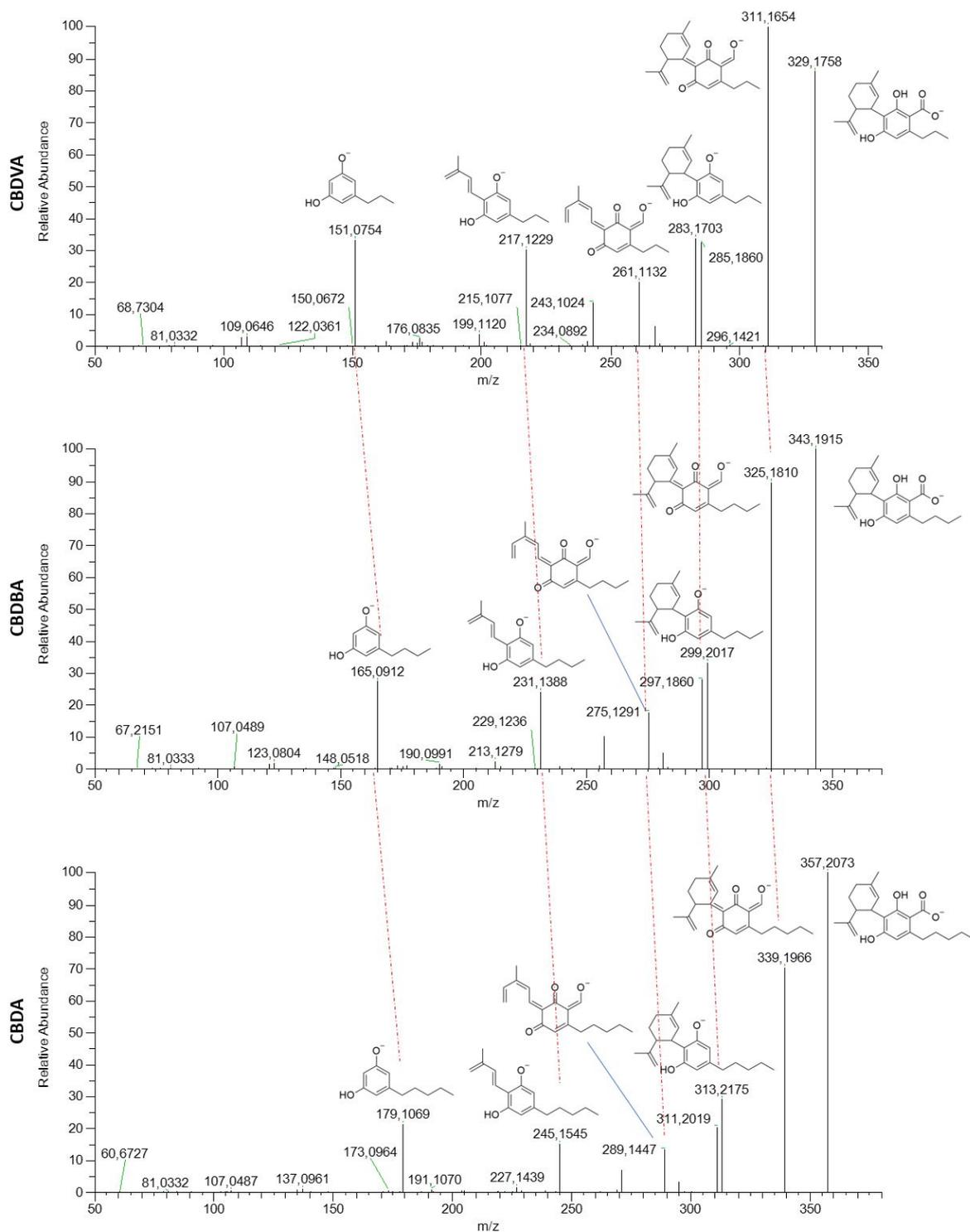
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TABLE OF CONTENT

HRMS spectra of carboxylated and neutral cannabinoids in positive and negative mode (Figures S1-6)	...SI-2
NMR spectra of synthetic (-)-<i>trans</i>-Δ^9-THCB (Figures S7-11)	...SI-8
NMR spectra of extracted (-)-<i>trans</i>-Δ^9-THCB (Figures S12-16)	...SI-13
Superimposition of ¹H NMR and ¹³C NMR of extracted and synthetic (-)-<i>trans</i>-Δ^9-THCB (Figure S17)	...SI-19
Circular Dichroism (CD) spectra of isolated and synthesized THCB (Figure S18)	...SI-19

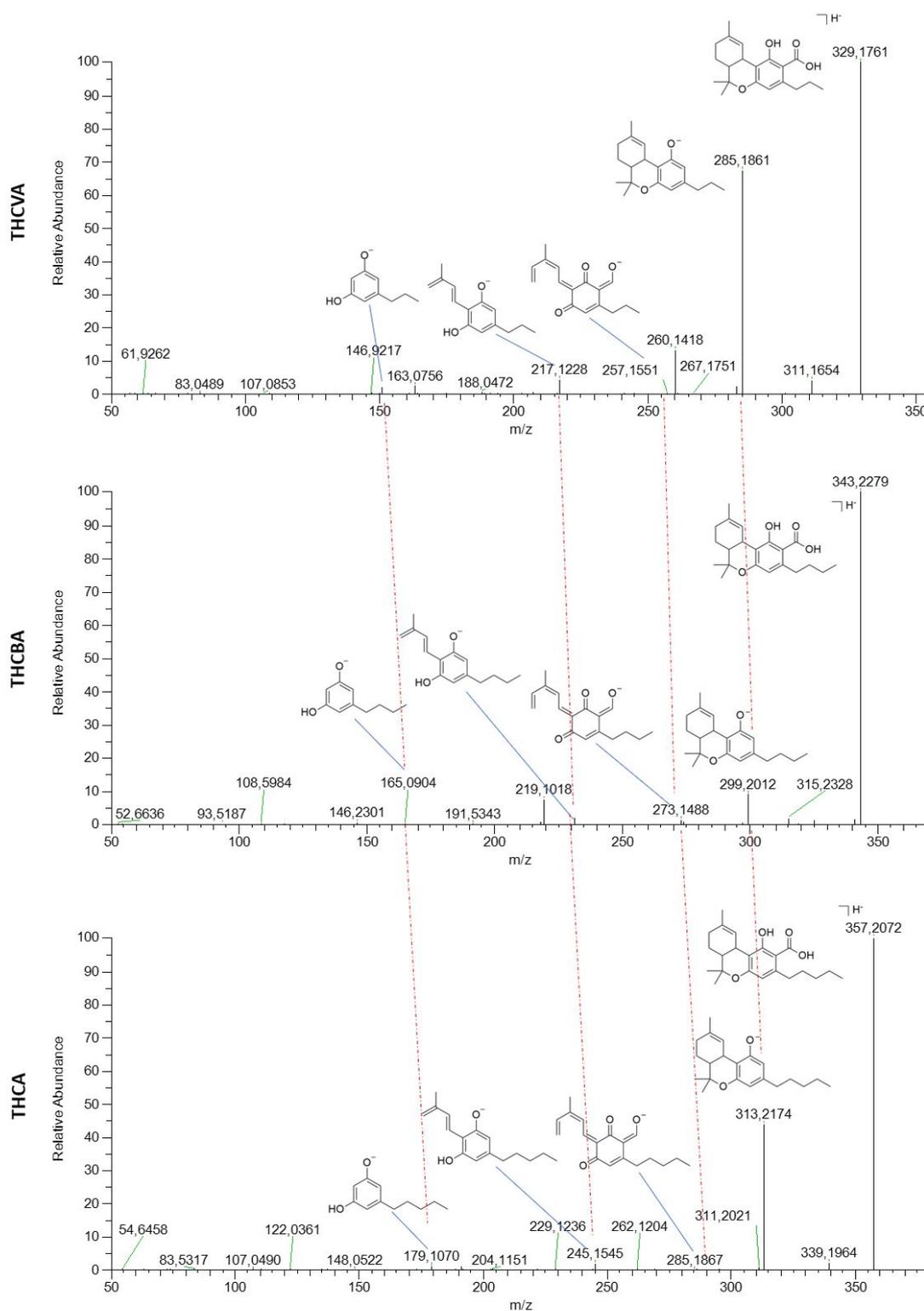
HRMS spectra of carboxylated and neutral cannabinoids

Figure S1. HRMS spectra of CBDA, CBDBA and CBDVA in negative ionization mode. A putative structure is given for each fragment. Dotted red lines indicate correspondence of fragments between pentyl (m/z), butyl ($m/z-\text{CH}_3$) and propyl ($m/z-\text{C}_2\text{H}_5$) forms.



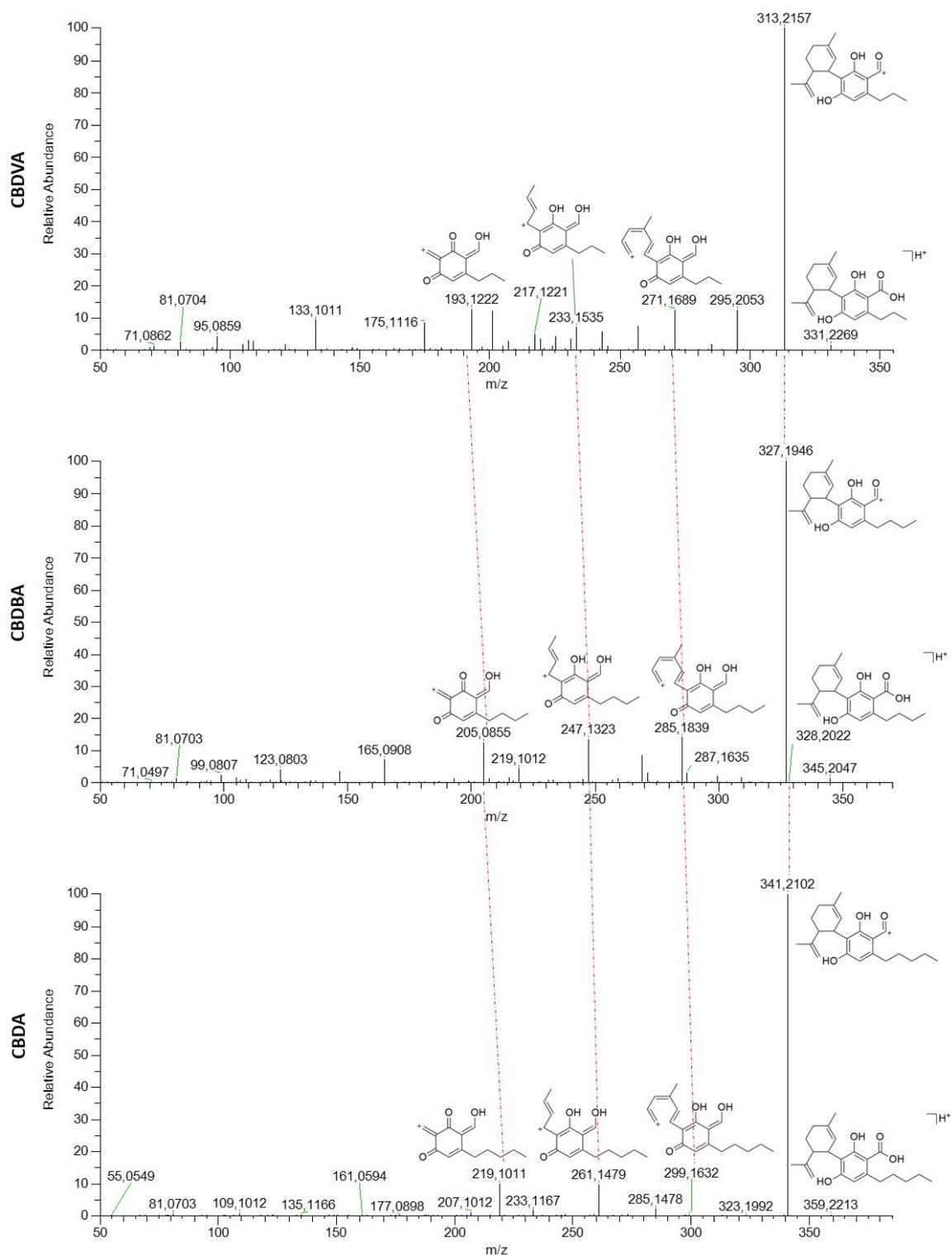
SUPPORTING INFORMATION

Figure S2. HRMS spectra of THCA, THCBA and THCVA in negative ionization mode. A putative structure is given for each fragment. Dotted red lines indicate correspondence of fragments between pentyl (m/z), butyl (m/z -CH₃) and propyl (m/z -C₂H₅) forms.



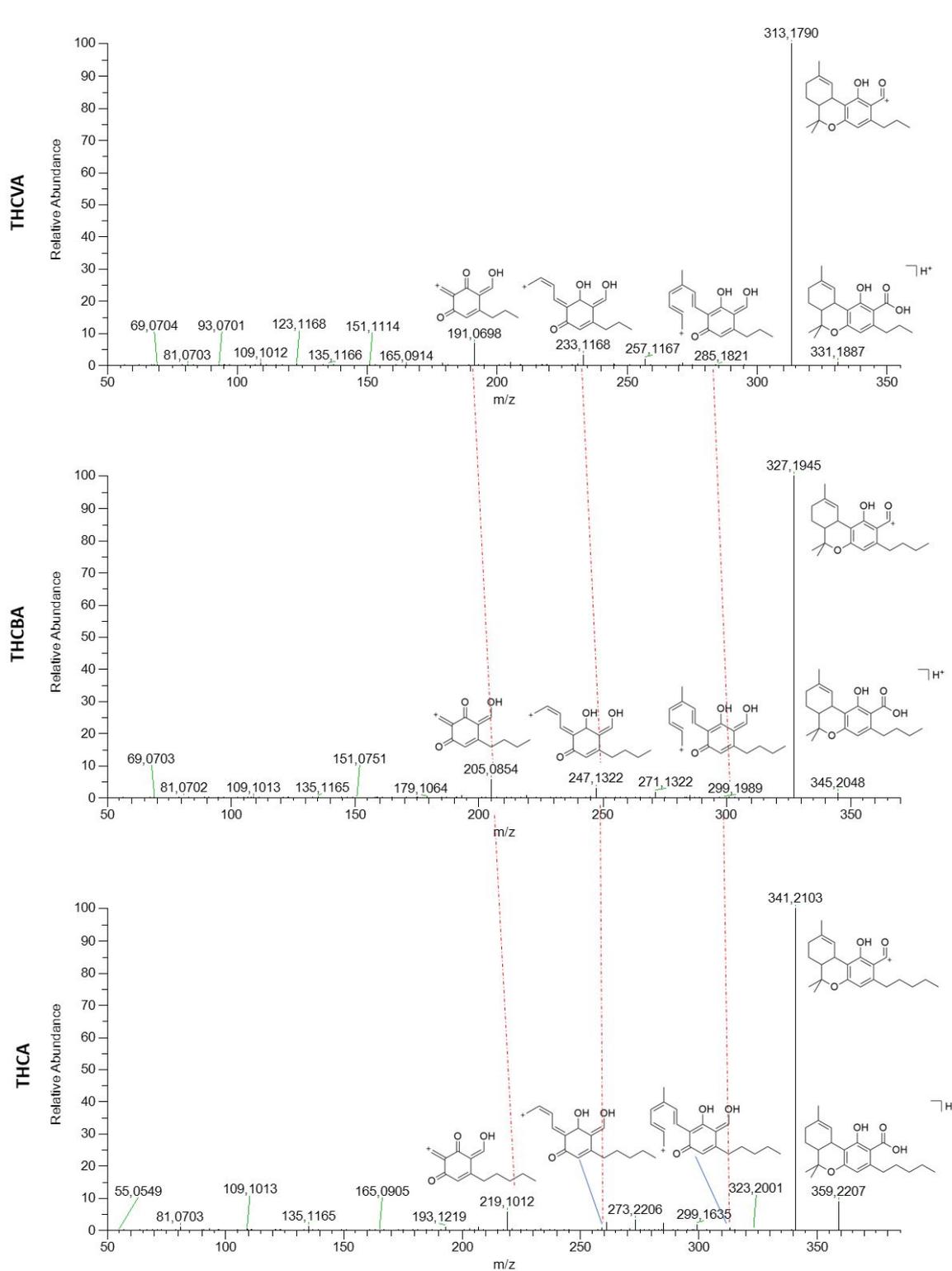
SUPPORTING INFORMATION

Figure S3. HRMS spectra of CBDA, CBDDBA and CBDVA in positive ionization mode. A putative structure is given for each fragment. Dotted red lines indicate correspondence of fragments between pentyl (m/z), butyl (m/z -CH₃) and propyl (m/z -C₂H₅) forms.



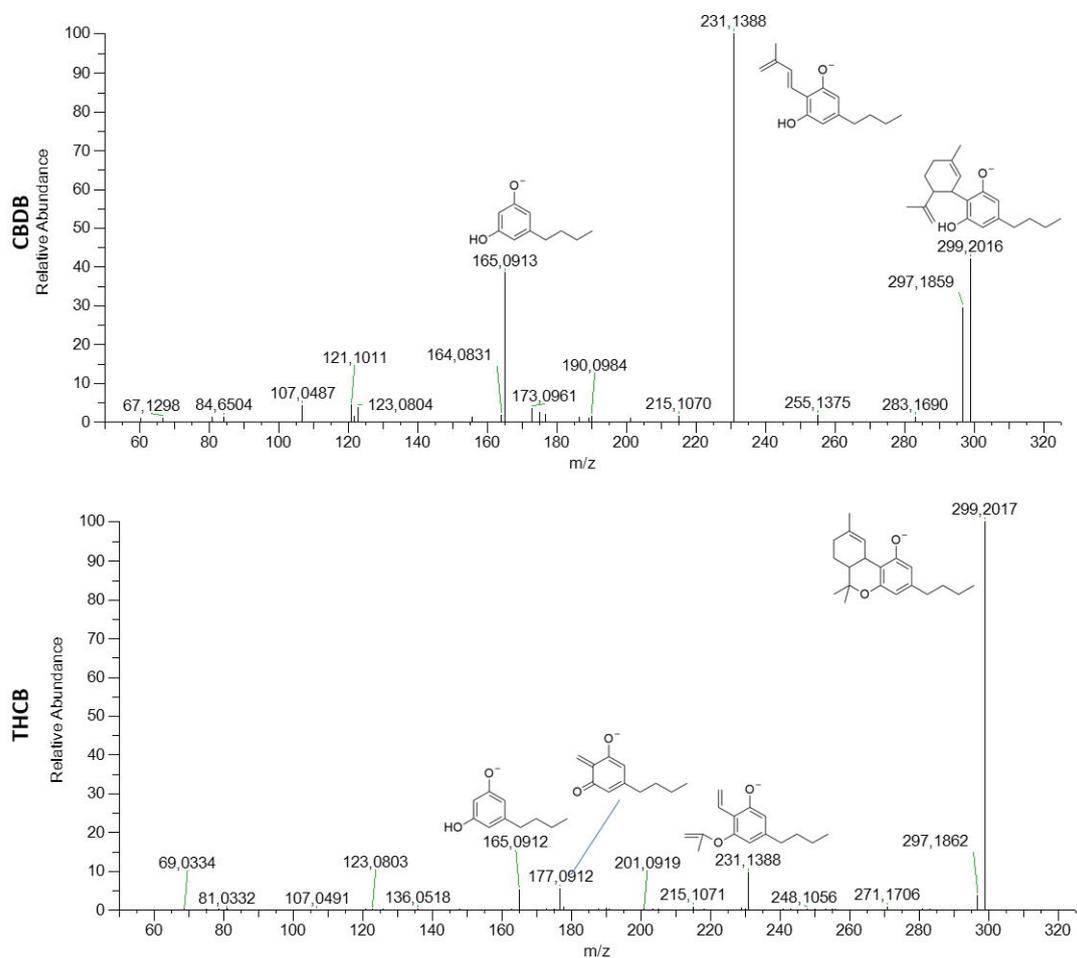
SUPPORTING INFORMATION

Figure S4. HRMS spectra of THCA, THCBA and THCVA in positive ionization mode. A putative structure is given for each fragment. Dotted red lines indicate correspondence of fragments between pentyl (m/z), butyl ($m/z-CH_3$) and propyl ($m/z-C_2H_5$) forms.



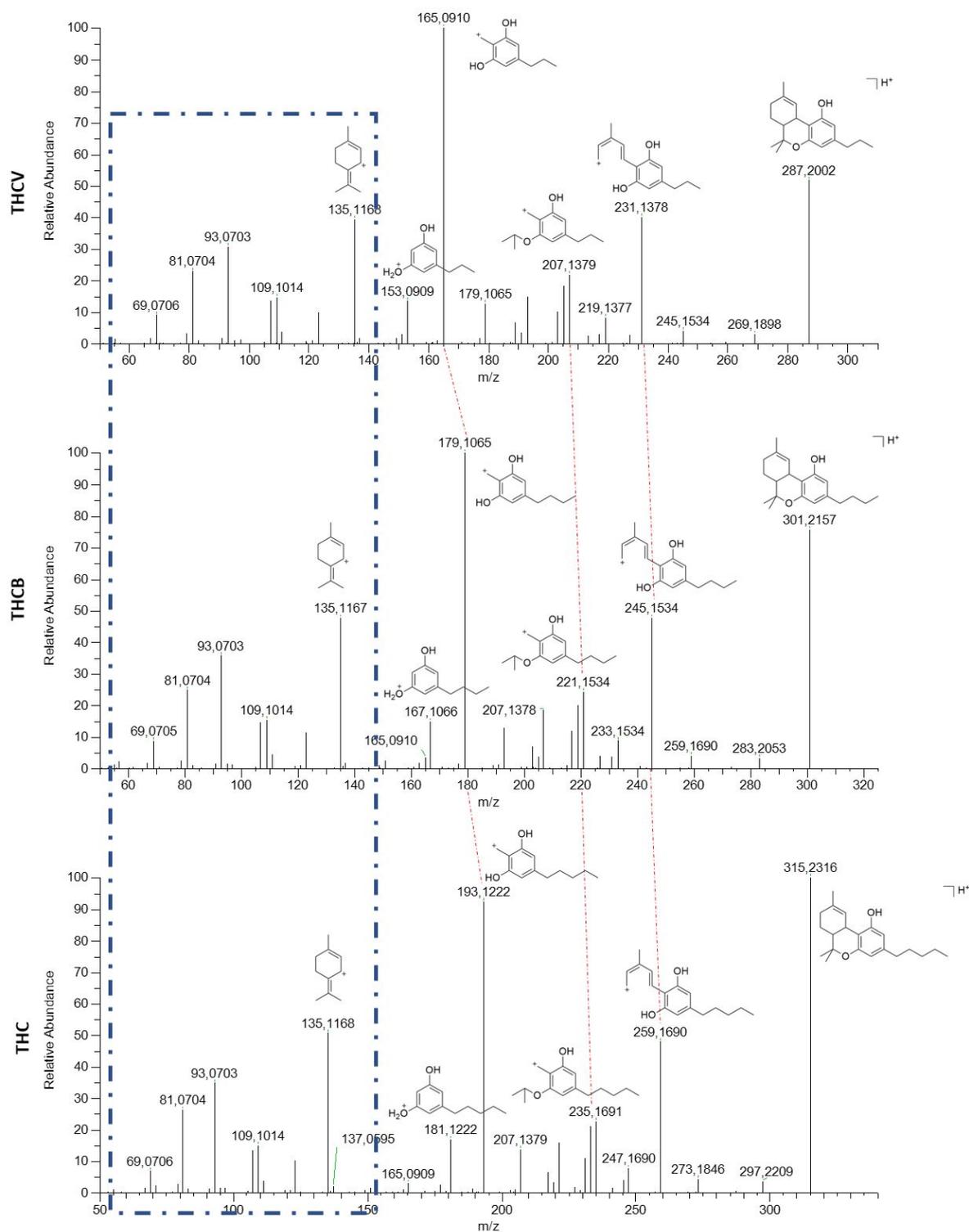
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Figure S5. Comparison of CBDB (top) and Δ^9 -THCB (bottom) HRMS spectrum in negative ionization mode. A putative structure is given for each fragment.



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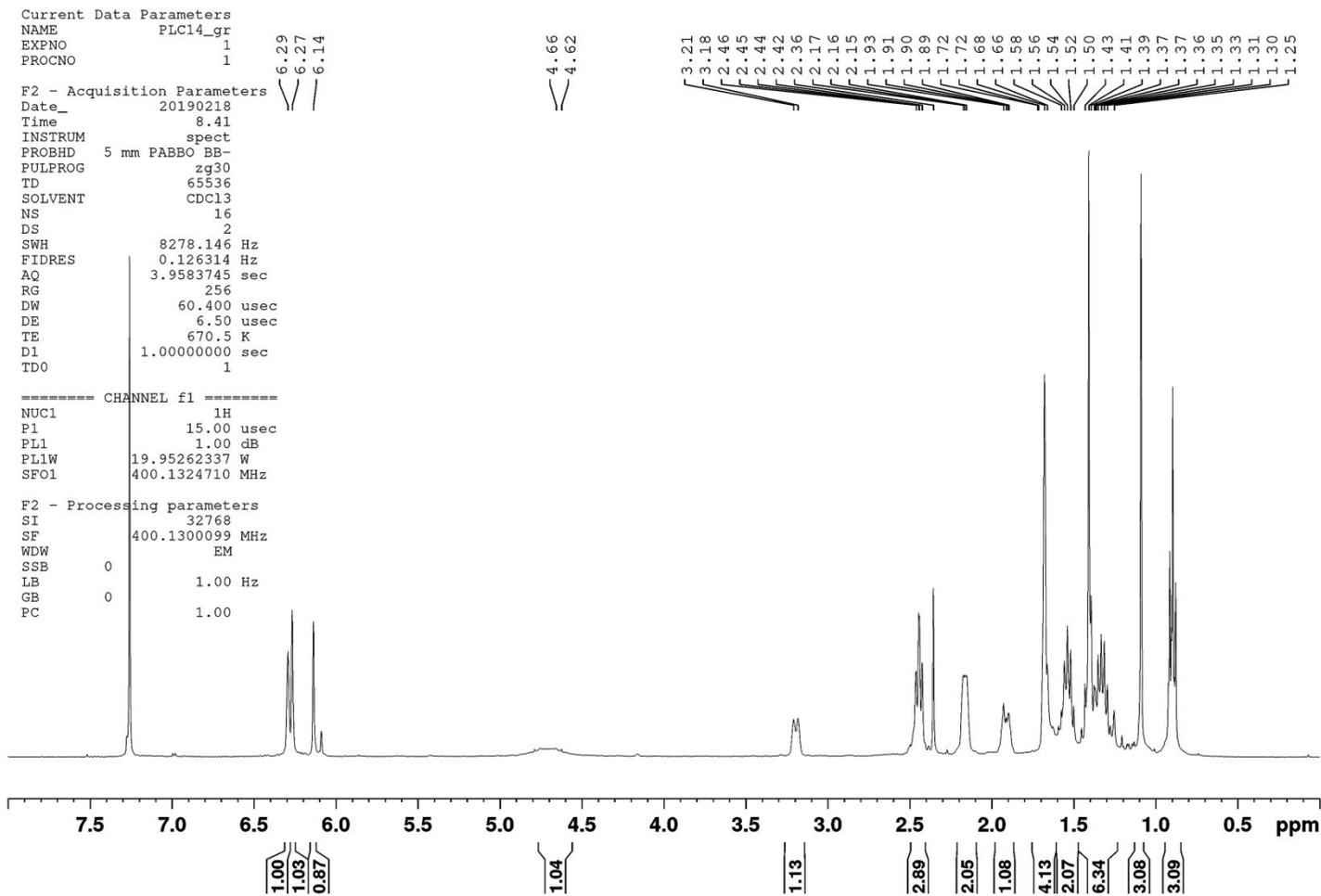
Figure S6. HRMS spectra of Δ^9 -THC, Δ^9 -THCB and Δ^9 -THCV in positive ionization mode. A putative structure is given for each fragment. Dotted red lines indicate correspondence of fragments between pentyl (m/z), butyl (m/z -CH₃) and propyl (m/z -C₂H₅) forms.



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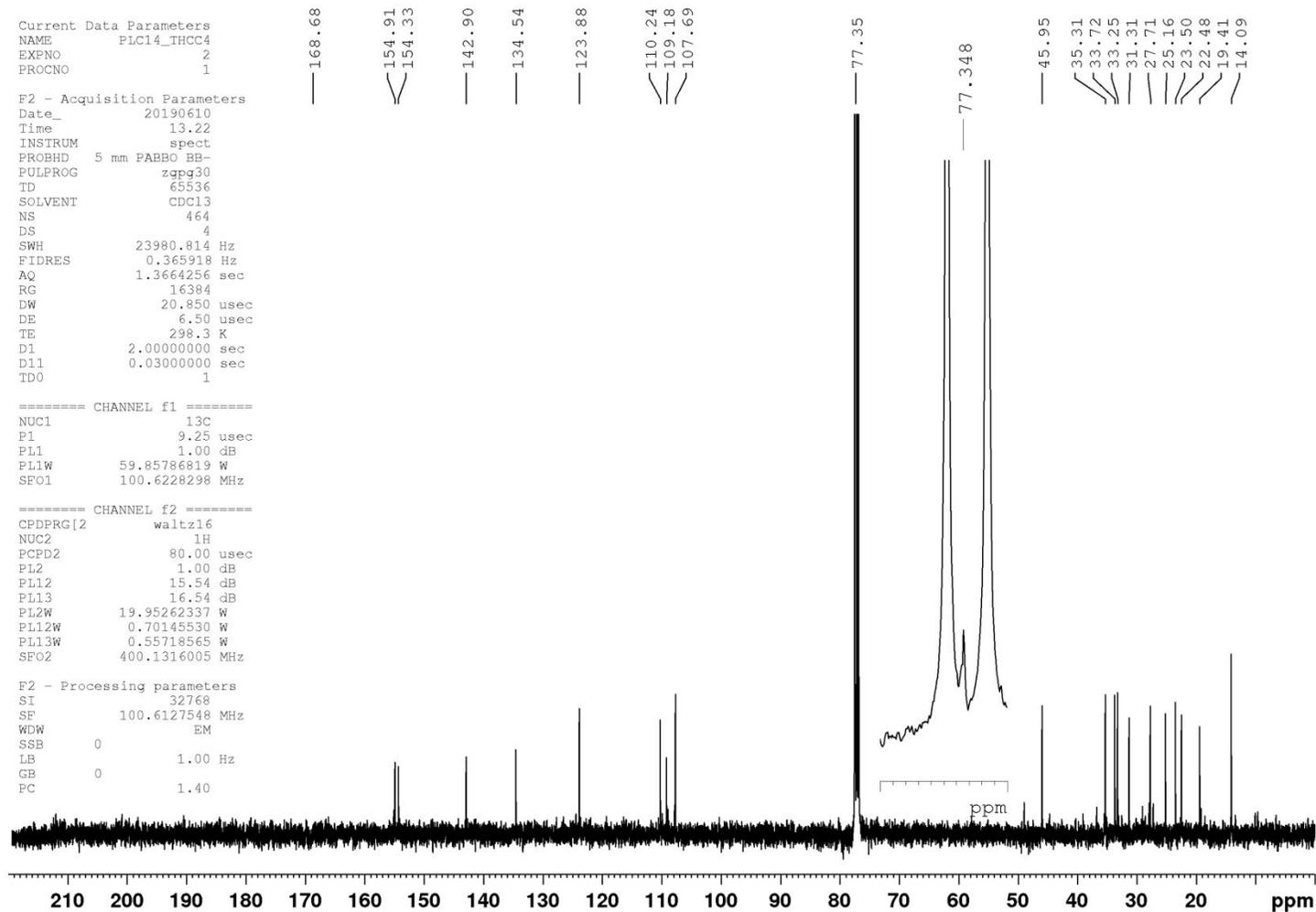
NMR spectra of synthetic (-)-*trans*- Δ^9 -THCB

Figure S7



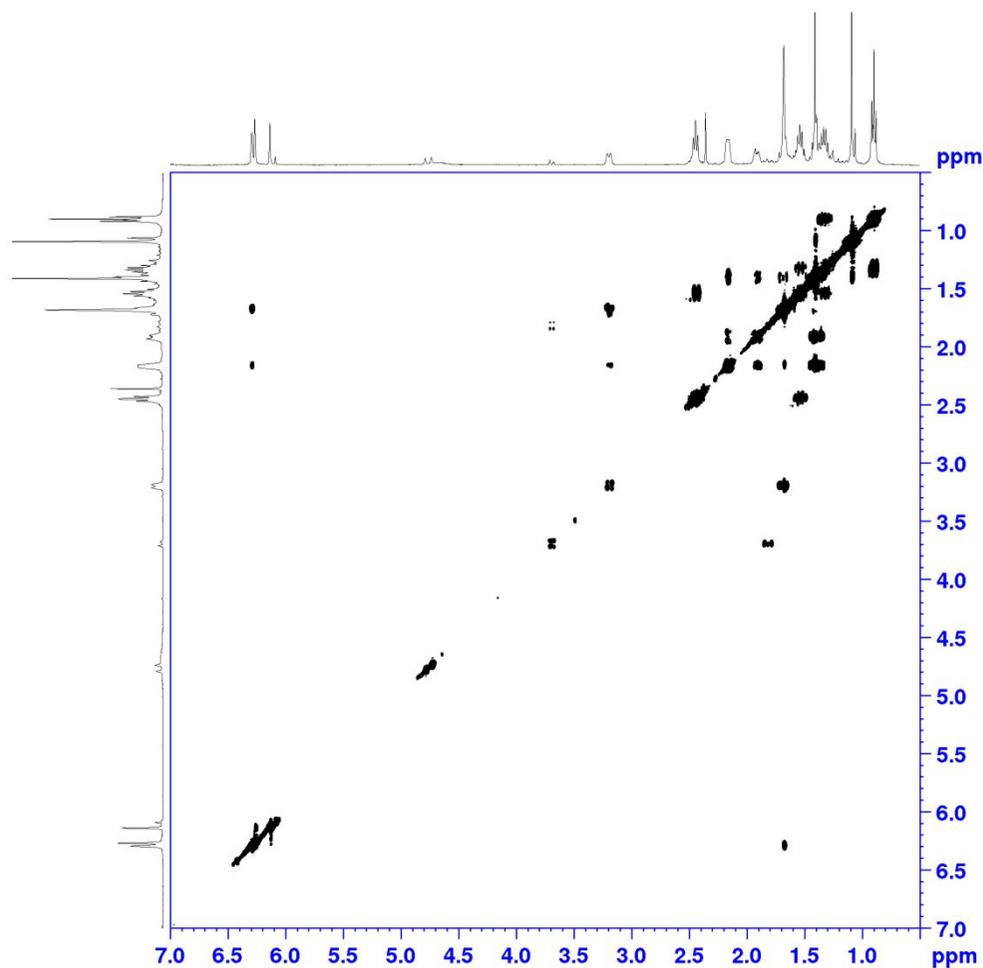
SUPPORTING INFORMATION

Figure S8



SUPPORTING INFORMATION

Figure S9



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PROCNO    1

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TD         2048
SOLVENT   CDCl3
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TE         297.8 K
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INO        0.00018720 sec

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P1         15.00 usec
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SFO1       400.1324057 MHz

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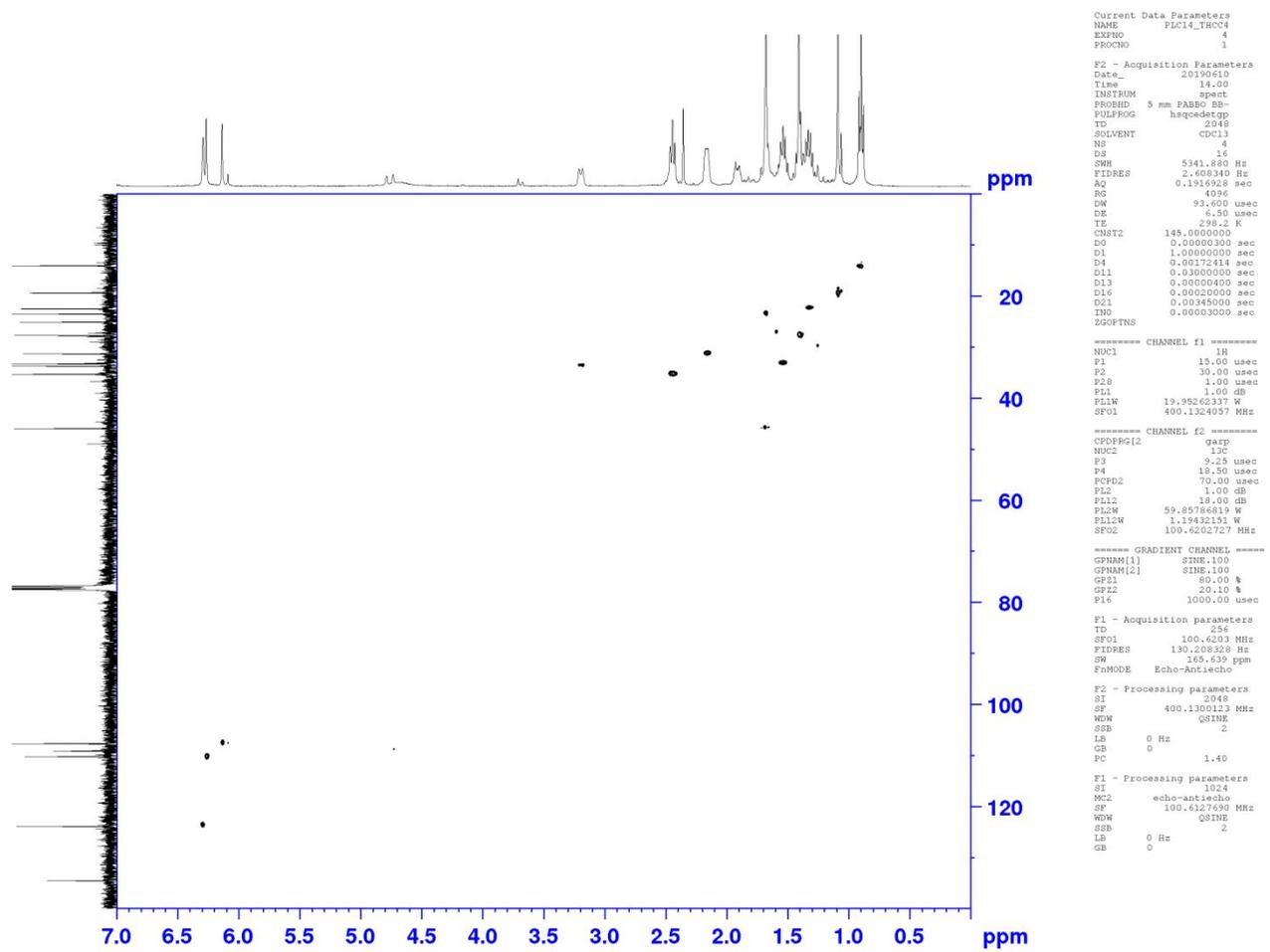
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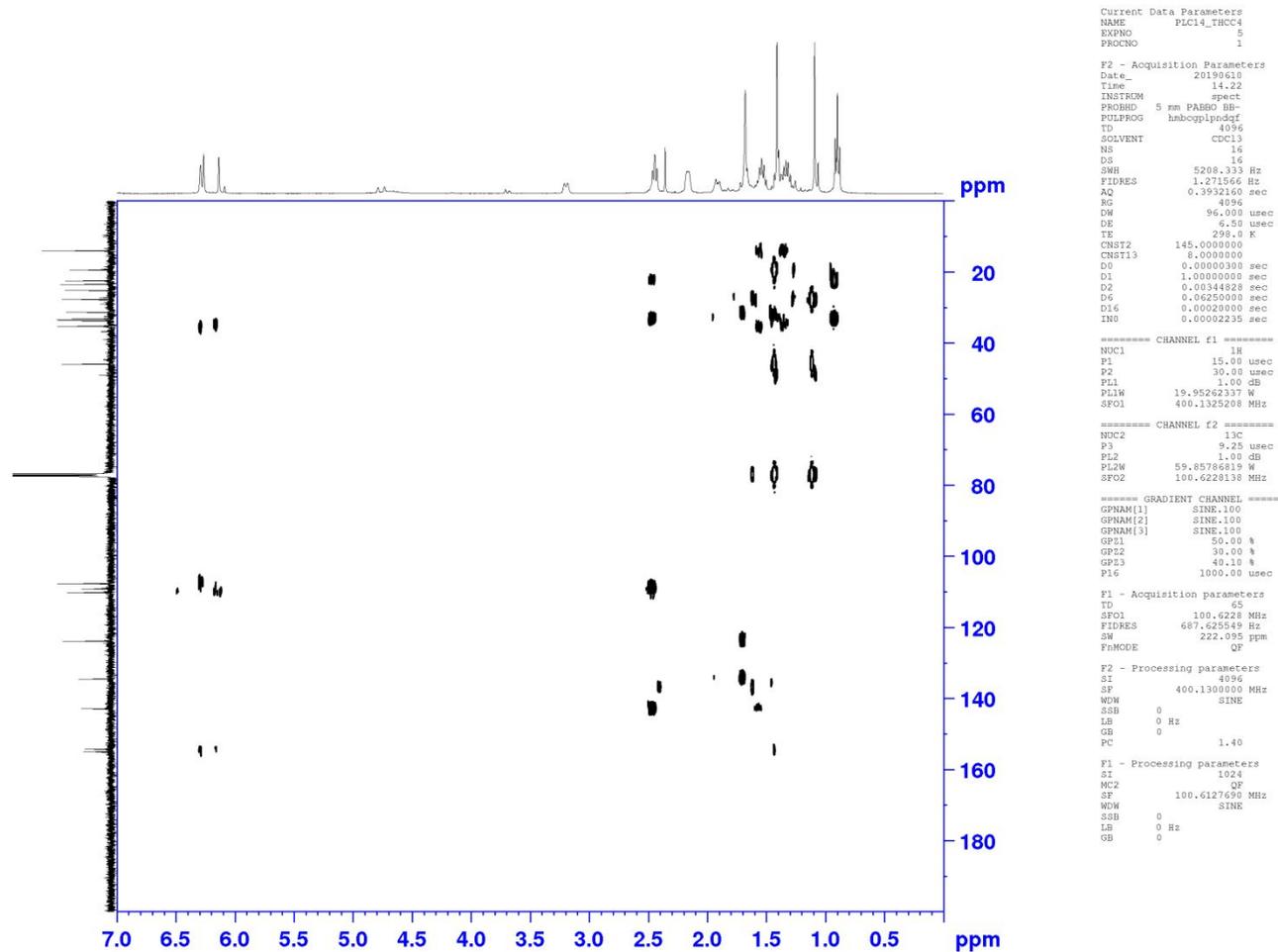
SUPPORTING INFORMATION

Figure S10



SUPPORTING INFORMATION

Figure S11



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PROBHD   5 mm PABBO BB-
PULPROG  lmbogp1pdgdf
TD       4096
SOLVENT  CDCl3
NS       16
DS       16
SWH      5208.333 Hz
FIDRES   1.271566 Hz
AQ       0.3932160 sec
RG       4096
DW       96.000 usec
DE       6.50 usec
TE       299.0 K
CNST2    145.0000000
CNST13   8.0000000
D0       0.0000000 sec
D1       1.0000000 sec
D2       0.00344828 sec
D6       0.00250000 sec
D16      0.00020000 sec
IN0      0.00002235 sec

===== CHANNEL f1 =====
NUC1     1H
P1       15.00 usec
P2       30.00 usec
PL1      1.00 dB
PL1W     19.95262337 W
SFO1     400.1325208 MHz

===== CHANNEL f2 =====
NUC2     13C
P3       9.25 usec
P4       1.00 dB
PL2W     59.85786819 W
SFO2     100.6228138 MHz

===== GRADIENT CHANNEL =====
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GPNAM[2] SINE.100
GPNAM[3] SINE.100
GP21     50.00 %
GP22     30.00 %
GP23     40.10 %
P16      1000.00 usec

F1 - Acquisition parameters
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SFO1     100.6228 MHz
FIDRES   687.625549 Hz
SW       222.095 ppm
F2MODE   QF

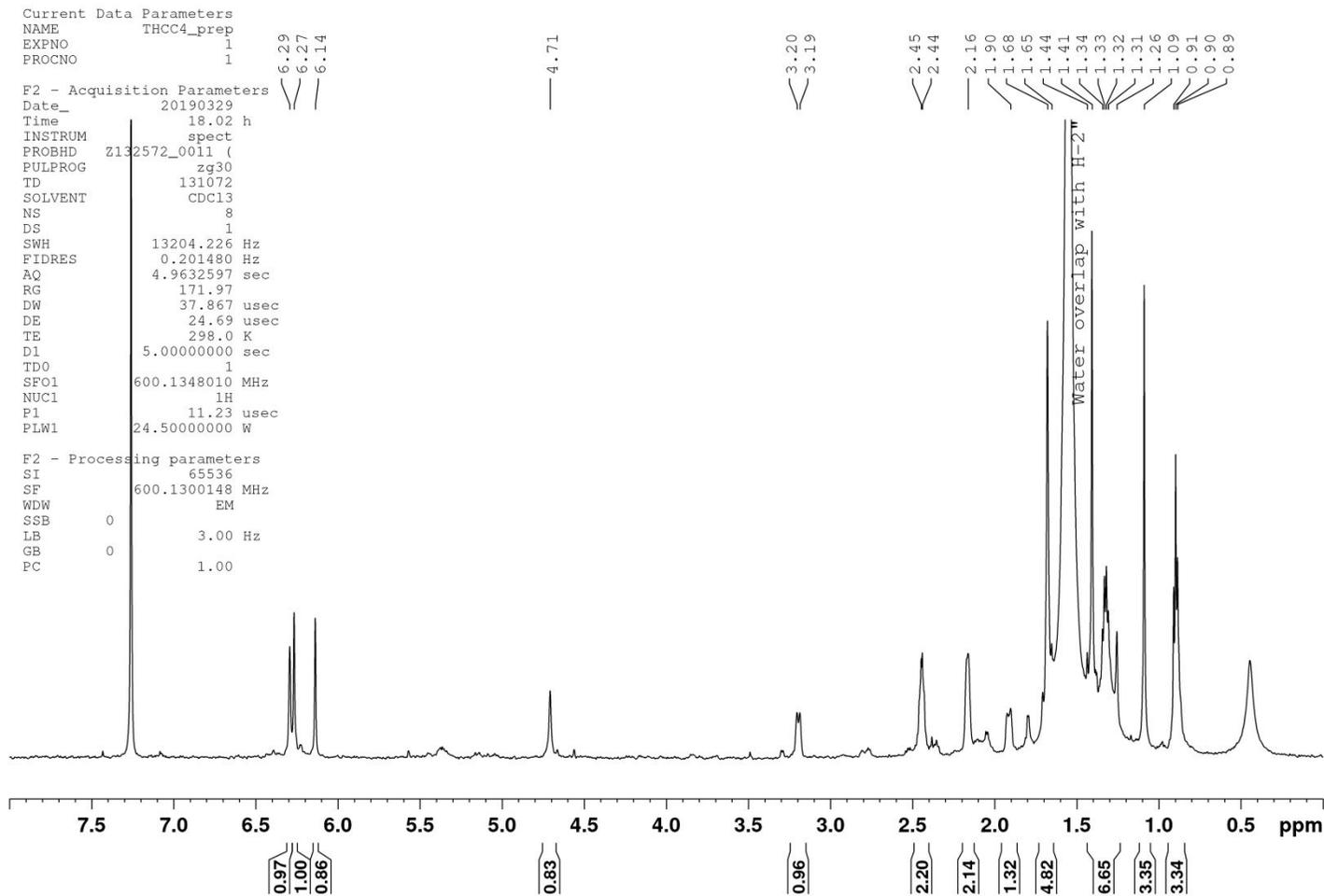
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PC       1.40

F1 - Processing parameters
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SUPPORTING INFORMATION

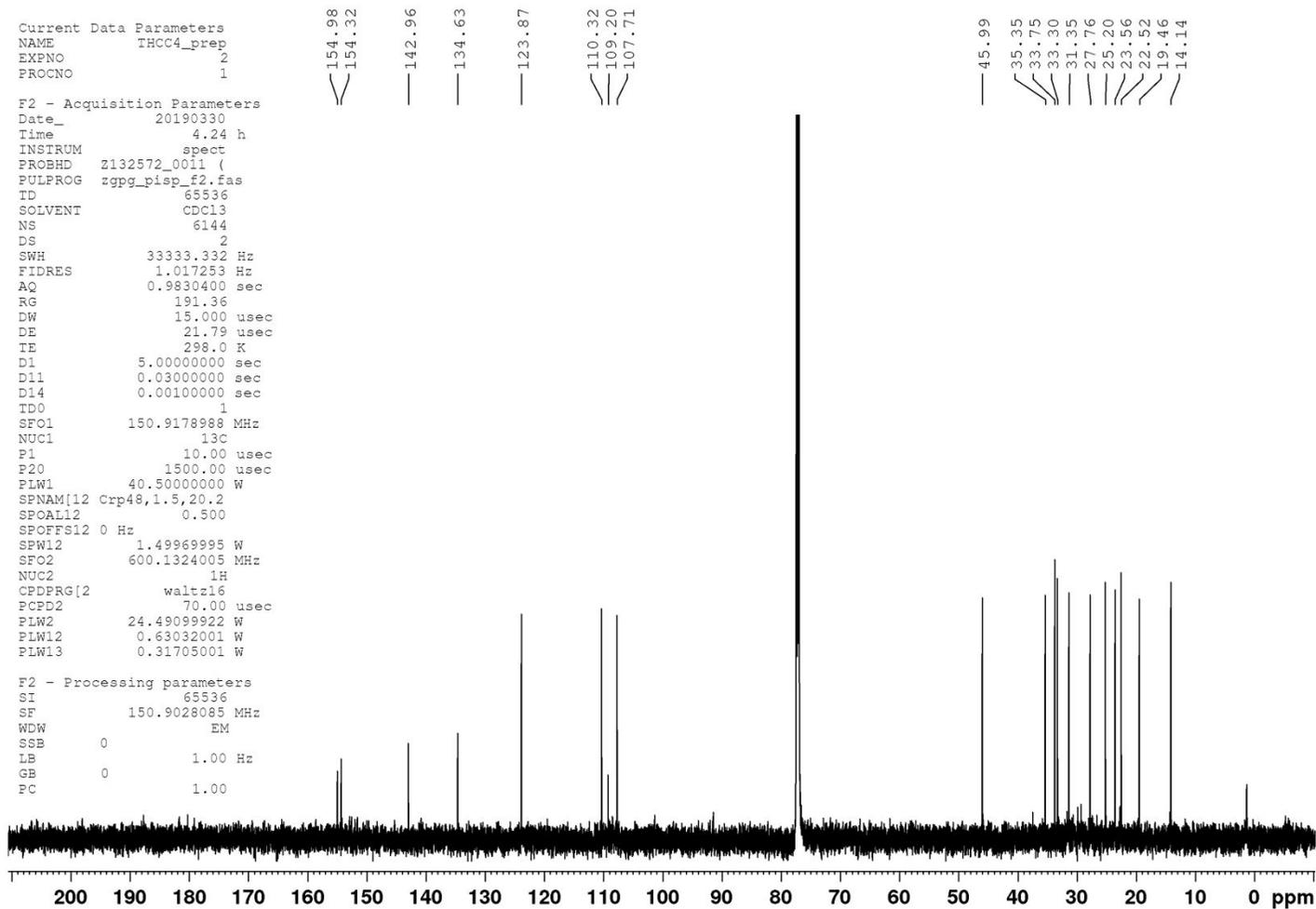
NMR spectra of extracted (-)-*trans*- Δ^9 -THCB

Figure S12



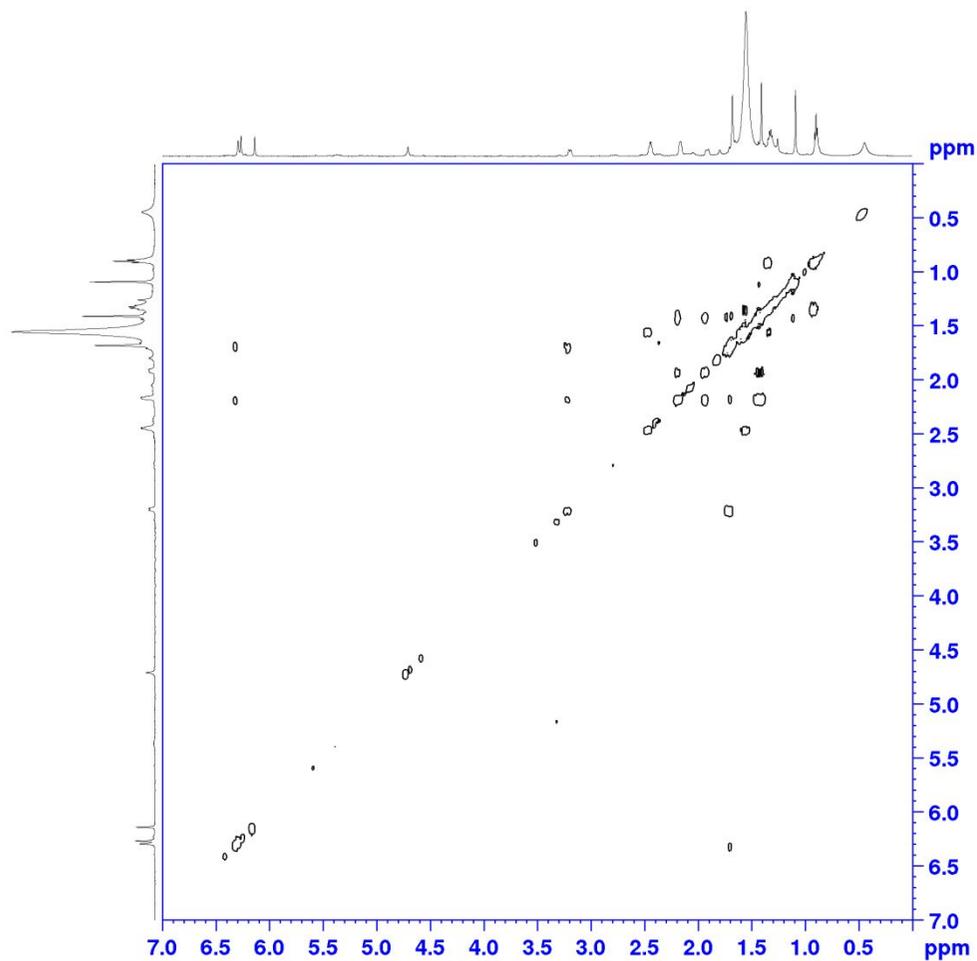
SUPPORTING INFORMATION

Figure S13



SUPPORTING INFORMATION

Figure S14



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Current Data Parameters
NAME      THCC4_prep
EXPNO     3
PROCNO    1

F2 - Acquisition Parameters
Date_     20190330
Time      4.24 h
INSTRUM   spect
PROBHD    Z132572_0011 (
PULPROG   cosyppppf
TD         2048
SOLVENT   CDCl3
NS         4
DS         8
SWH        7812.500 Hz
FIDRES     7.629395 Hz
AQ         0.1310720 sec
RG         54.87
DW         64.000 usec
DE         20.00 usec
TE         298.0 K
D0         0.00000300 sec
D1         1.00000000 sec
D11        0.03000000 sec
D12        0.00002000 sec
D13        0.00000400 sec
D16        0.00020000 sec
INO        0.00012800 sec
TDAV      1
SF01      600.1324005 MHz
NUC1       1H
P0         11.23 usec
P1         11.23 usec
P17        2500.00 usec
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PLW10      4.94169998 W
GPNAM[1]   SMSQ10.100
GPZ1       10.00 %
P16        1000.00 usec

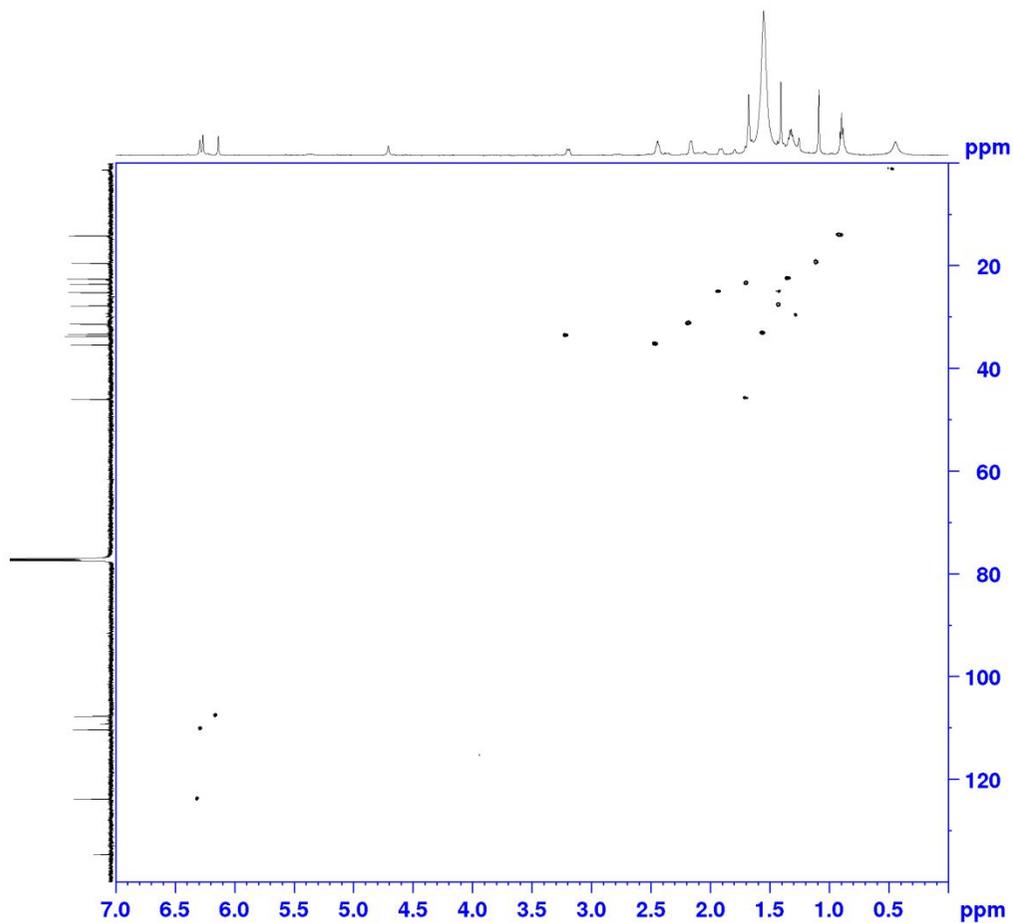
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SW         13.018 ppm
FhMODE     QF

F2 - Processing parameters
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SF         600.1300000 MHz
WDW        QSINE
SSB        0
LB         0 Hz
GB         0
PC         1.00

F1 - Processing parameters
SI         1024
MC2        QF
SF         600.1300000 MHz
WDW        QSINE
SSB        0
LB         0 Hz
GB         0
    
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SUPPORTING INFORMATION

Figure S15



```

Current Data Parameters
NAME      THCC4_prep
EXPNO    4
PROCNO   1

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Date_    20190330
Time     4.45 h
INSTRUM  spect
PROBHD   2132572_0011 (
PULPROG  hsqcetppp3
TD       2048
SOLVENT  CDCl3
NS       8
DS       16
SWH      7612.500 Hz
FIDRES   7.629395 Hz
AQ       0.1310720 sec
RG       191.36
DW       64.000 usec
DE       20.00 usec
TE       298.0 K
CNS12    145.0000000
D0       0.00000000 sec
D1       0.50000000 sec
D4       0.00172414 sec
D11      0.03000000 sec
D16      0.00020000 sec
D21      0.00345000 sec
DNO      0.00001840 sec
TDav     1
SFO1     600.1324005 MHz
NUC1     1H
P1       11.23 usec
P2       22.46 usec
P28      0 usec
PLW1     24.49099922 W
PFC2     150.9148808 MHz
NUC2     13C
CHFPFG(2)  gapp4
P3       10.00 usec
P14      500.00 usec
P21      1730.00 usec
PCPD2    60.00 usec
PLW0     0 W
PLW2     40.50000000 W
PLW12    1.12500000 W
SFXM[3]  Crp60,0.3,2.0,1
SFOAL1   0.500
SFOFFS3  0 Hz
SFXM3    6.18790007 W
SFXM[18] Crp60_xfilr.2
SFOAL18  0.500
SFOFFS18 0 Hz
SFXM18   1.78840005 W
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GFC2     20.10 %
P16      1000.00 usec

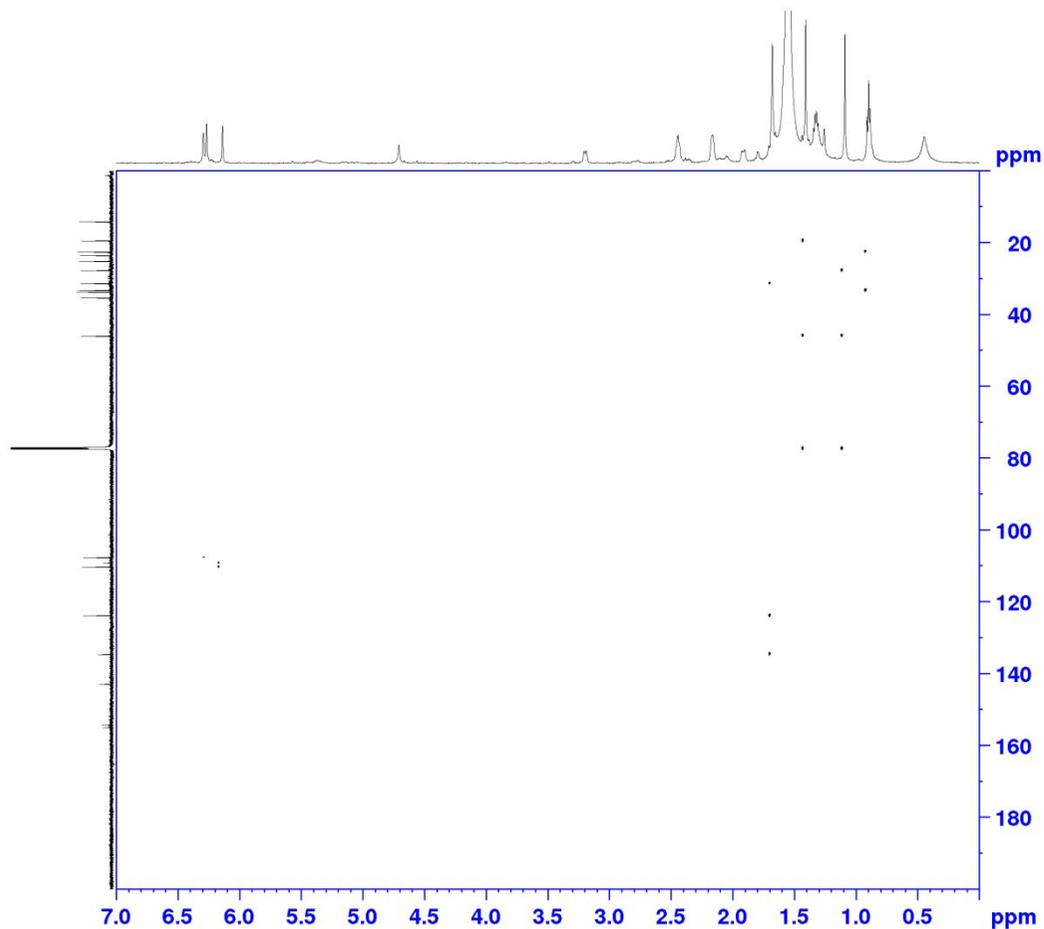
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F1M0DE   Echo-Antiecho

F2 - Processing parameters
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SF       600.1300000 MHz
WDW      QSIINE
SSB      2
LB       0 Hz
GB       0
PC       1.00

F1 - Processing parameters
SI       1024
MCE      echo-antiecho
SF       150.9028085 MHz
WDW      QSIINE
SSB      2
LB       0 Hz
GB       0
    
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SUPPORTING INFORMATION

Figure S16



```

Current Data Parameters
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EXEND    6
PROCNO    1

F2 - Acquisition Parameters
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Time      5.30 h
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PULPROG   hmbcctegp13nd
TD         4096
SOLVENT    CDCl3
NS         16
DS         16
SWH        7812.500 Hz
FIDRES     3.814697 Hz
AQ         0.2621440 sec
RG         191.36
DW         64.000 usec
DE         20.000 usec
TE         298.0 K
CNS15     120.000000
CNS17     170.000000
CNS13     8.000000
D0         0.0000000 sec
D1         0.5000000 sec
D6         0.06250000 sec
D16        0.00020000 sec
IN0        0.00001510 sec
TD0v      1
SFO1       600.1324005 MHz
NUC1       1H
P1         11.23 usec
P2         22.46 usec
PLW1       24.49099922 W
SFO2       150.9178988 MHz
NUC2       13C
P3         10.00 usec
P4         2000.00 usec
PLW2       40.50000000 W
SPNAM[7]   Crp60comp.4
SFOAL[7]   0.500
SFOFF[7]   0 Hz
SFW       6.18790007 W
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GPF1       80.00 %
GPNAM[3]   SMSQ10.100
GPF3       14.00 %
GPNAM[4]   SMSQ10.100
GPF4       -8.00 %
GPNAM[5]   SMSQ10.100
GPF5       -4.00 %
GPNAM[6]   SMSQ10.100
GPF6       2.00 %
P16        1000.00 usec

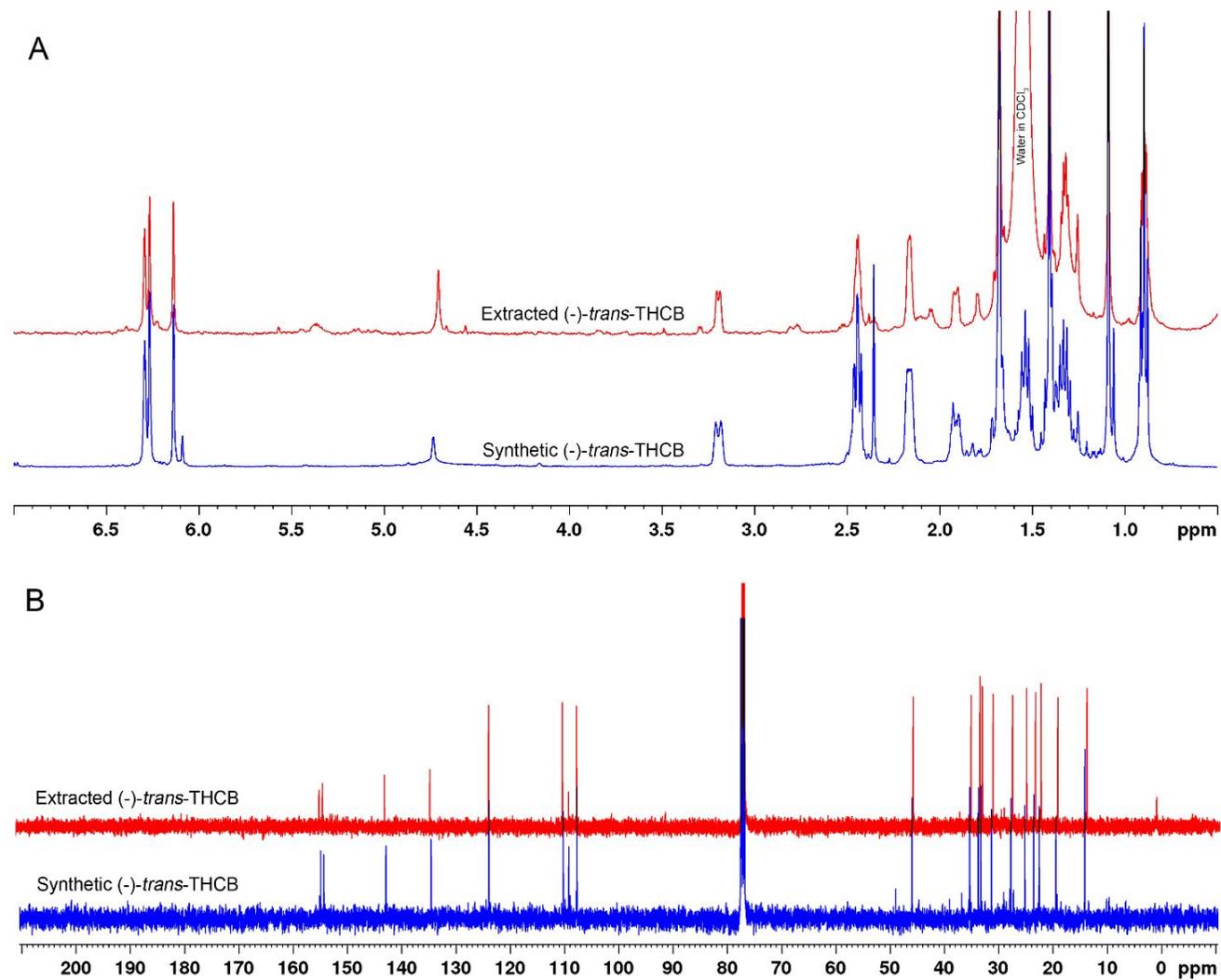
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SSB        4
LB         0 Hz
GB         0
PC         1.00

F1 - Processing parameters
SI         1024
MC2        echo-antiecho
SF         150.9028085 MHz
WDW        QSINE
SSB        2
LB         0 Hz
GB         0
    
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SUPPORTING INFORMATION

Figure S17. Superimposition of ^1H NMR (A) and ^{13}C NMR (B) of extracted (-)-*trans*- Δ^9 -THCB (red spectra) and synthetic (-)-*trans*- Δ^9 -THCB (blue spectra).



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Figure S18. Superimposition of the Circular Dichroism (CD) spectra of isolated (green) and synthesized (blue) Δ^9 -THCB, in acetonitrile at 10 $\mu\text{g/mL}$.

